Vol 4

No.

DOI: 10.21767/2572-5459.100051

# Simple Methods to Estimate Total Body Water in Live Animals Using Antipyrine with Detection of Heat Adaptability

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Received date: August 26, 2018; Accepted date: January 3, 2019; Published date: January 7, 2019

Citation: Habeeb AAM (2019) Simple Methods to Estimate Total Body Water in Live Animals Using Antipyrine with Detection of Heat Adaptability. J Anim Res Nutr 4: 1: 1.

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### **Abstract**

Estimating body water content in live animal using Antipyrine substance as a new and modified technique was the objective of this research. Body water was estimated in five calves with average body weight of 183.6 ± 30.3 kg using conventional method (extrapolation technique) and in the same time by modified method. The averages of total body water in five calves were 135.8  $\pm$  11.8 and 132.3  $\pm$ 11.35 liters by convention and modified technique, respectively, without significant difference between the two techniques. The corresponding summations of body water in all calves were 679.2 and 661.4 liters. The accuracy of modified technique was 97.4 % as compared with convention method.

It is concluded that estimate body water using Antipyrine by new method is easily, accurate and quickly technique and more reliable. Animals when exposed to high ambient temperature during hot summer season, water intake increase and consequently body water content increases with different percentage according animal response to stressful conditions. The percentage change in body water or body solids (Live body weight- body water) contents in each animal may be used for evaluation the animal's adaptability to heat stress.

**Keywords:** Body water; Antipyrine; Calves; Body solids

### Introduction

The total body water pool is all the water in the animal including the alimentary tract, which has a large volume, particularly in ruminants. Body water is the water content of an animal body that is contained in the tissues, the blood, the bones and elsewhere. About 99% of all molecules in the body are water, which forms about 70% of the body weight of a tropical ruminant. About 45% of bodyweight is intracellular water and 25% is extracellular, divided between plasma (5%) and interstitial fluids (20%) [1].

Body water in live animals is important for research whether the research involves nutrition, physiology, genetic, disease and meat production. Estimation of body water in animals using slaughter and chemical analysis of the whole bodies organs is a tedious processing, time-consuming and expensive operation [2]. Besides that the high cost of animal analysis has created an interest in indirect methods of estimating body water. This indirect method or in vivo also can provide repeated estimates of body water for same animal whereas slaughter and chemical analysis obviously can only be done once [3].

Moreover, body weight of animal alone provides a poor index of the metabolically active tissue due to that body weight is including body solids and body water, consequently using live body weight for estimating body weight gain of animals is a misleading index of growth performance, since it may be due to the increase in water retention and not to the increase in body protein and fat. In other words, a unit of body weight gain in one animal may be due to the increase in body water at the expense of body tissue loss, while in the other animal, may be due to the increase in body solids [4].

Most methods for measuring the total water content of the body in vivo have been based on the degree of dilution of a foreign substance or radioisotope after its intravenous injection. This substance or radioisotope should possess the following characteristics: even and rapid distribution throughout body water; non-toxicity in required doses; slow transformation in, and excretion from, the body; accurate and convenient estimation of slow its concentration in the plasma. Application of the isotopes materials using tritium or deuterium for measuring total body water after counting ß-activity in Liquid Scintillation Counting was carried out by different authors [5-10].

There are many disadvantages in using isotopes in estimation of total body water in live animals. This technique needs special instrument like liquid scintillation counting with liquid scientillator solution to counting ß-activity. In addition, these animals after injection with radioisotopes must be culled. Therefore, antipyrine (ANP) as nonradioactive substance may be used in estimation of body water in live animals.

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Measuring the total body water of the animal *in vivo* by ANP has been developed by Brodie et al. [11]. This conventional method of Brodie involves the use of ANP (l-phenyl-2, 3-dimethylpyrazolone-5-one) for estimating total body water by injection 1 gm/100 kg body weight of antipyrine in distilled water intravenously from calibrated syringe and five blood samples are withdrawn at 2, 3, 4 and 5 hours subsequently and protein precipitation for plasma. ANP is measured in filtrate from the ultraviolet absorption of 4-nitroso-antipyrine by the addition of sodium nitrite and sulphuric acid to plasma filtrate.]

The plasma concentration at zero time (the concentration at the time of injection) by plotting the plasma levels on semilogarithmic paper and extrapolating the straight portion of the time-concentration curve back to the time of injection by the method of least squares (extrapolation technique).

Estimating body water content in live animal using ANP by single blood sample at ½ hour after ANP injection as modified technique and comparison between the two methods for estimating body water in five calves was the objective of this research. In addition, how theoretically using total body water or total body solids in live animals for evaluation the animal's adaptability to heat stress was the second objective of this study.

## **Materials and Methods**

### **Location and ethics**

The experimental work was carried out in Bovine Farm of Biological Application Department, Radioisotopes Applications Division, Nuclear Research Centre, Atomic Energy Authority, at Inshas, Egypt (latitude 31º 12' N to 22 º 2' N, longitude 25 º 53' E to 35º 53' E).

This work was reviewed and approved by the Animal Care and Welfare Committee of Zagazig University, Egypt (ANWD-206). These ethics contain relevant information on the Endeavour to reduce animal suffering and adherence to best practices in veterinary care according to the International Council for Laboratory Animal Science (ICLAS) guidelines. Experimental animals were also cared using husbandry guidelines derived

from Egyptian Atomic Energy Authority standard operating procedures.

# Estimation of body water using antipyrine (ANP) using modified method

Injection dose of ANP: Standard dose is 1 gram ANP each 100 kg live body weight (LBW). Any human or animal weighted 100 kg need to 1 gram ANP for injection. The standard dose is 1 g/100 kg live body weight because assume TBW percentage = 50% i.e. 50 liter # 1 g/50 liter  $\rightarrow$ 1000000 µg ANP/50000 ml i.e. 100 µg/5 ml  $\rightarrow$  20 µg/ml  $\rightarrow$  inter reading of Spectrophotometer).

For preparation the injection dose of ANP, dissolve 20 gram ANP in physiological saline solution and complete the final solution to 100 ml  $\rightarrow$ 20 g/100 ml  $\rightarrow$ 1 g/5 ml. Each animal or person (weight 100 kg) inject with 5 ml (contains 1 g ANP) in the left jugular vein and one blood sample withdrawn from write jugular vein of animal after ½-1 hour from injection.

Chemical reagents required for antipyrine estimation: Zinc reagent solution: 10 g hydrolic zinc sulphate (ZnSO $_4$ .7H $_2$ O) was dissolved and 4 ml H $_2$ SO $_4$  (6 N) in distilled water and complete the solution with distilled water to reach 100 ml. This solution used in protein precipitation in the plasma samples. 10ZnSO $_4$ .7H $_2$ O + 4 ml H $_2$ SO $_4$  (6 N) distilled water $\rightarrow$ 100 ml zinc reagent solution.

**Sodium hydroxide (0.75 N):** 3 g recently dry sodium hydroxide was dissolved in distilled water and completes the solution with distilled water to reach 100 ml. 3 g NaOH dis.water $\rightarrow$ 100 ml Na OH (0.75 N)

**Sodium nitrite (0.2%):** Dissolve 0.1 g sodium nitrite was dissolved in distilled water and completes the solution with distilled water to reach 50 ml. 0.1 g Sodium nitrite dis.water→50 ml Sodium nitrite solutions (0.2%).

H<sub>2</sub>SO<sub>4</sub> acid with different normality as following in **Table 1**:

 $H_2SO_4$  concentration analar, Molecular weight = 98.08, Specific gravity = 1.84 i.e. 1 Liter = 1.84 kg # Concentration = 95-97% (96%).

**Table 1:** H<sub>2</sub>SO<sub>4</sub> acid with different normality and volume for ANP analysis

| H <sub>2</sub> SO <sub>4</sub> (6N)   | H <sub>2</sub> SO <sub>4</sub> (4N)  | H <sub>2</sub> SO <sub>4</sub> (0.07 N)   |  |
|---|--|---|--|
| 10 ml   | 10 ml  | 250 ml  |  |
| (6 x 98.08)/2=494.2 g/liter   | 4 x 98.08) /2=196.2 g /liter   | 0.07 x 98.08)/2=3.43 g /liter   |  |
| (494.2 g)/100=4.94 g/10 ml  | (196.2) / 100 =1.96 g /10 ml   | (3.43)/4=0.858 g/250 ml   |  |
| 4.94 g / 1.84 = 2.68 ml   | 1.96 g / 1.84 =1.06 ml   | 0.858 g/1.84=0.466 ml   |  |
| Conc.96%  | Conc.96%   | Conc. 96%   |  |
| (2.68 x 100)/96=2.79 ml   | (1.06 x 100) / 96 = 1.1 ml   | (0.446 x 100)/96= 0.49 ml   |  |
| Take 2.8 ml H <sub>2</sub> SO <sub>4</sub> con and complete with distilled water to reach the final volume 10 ml. | Take 1.1 ml H <sub>2</sub> SO <sub>4</sub> con. and complete with distilled water to reach the final volume 10 ml. | Take $\frac{1}{2}$ ml H $_2$ SO $_4$ con. and complete with distilled water to reach the final volume 250 ml. |  |
| →10 ml H <sub>2</sub> SO <sub>4</sub> (6 N)   | →10 ml H <sub>2</sub> SO <sub>4</sub> (4 N)  | →250 ml H <sub>2</sub> SO <sub>4</sub> (0.07 N)   |  |

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**Preparation of standard ANP:** One ml from injection dose (contains 400  $\mu$ g ANP) was put and complete the solution with H<sub>2</sub>SO<sub>4</sub> (0.07 N) to reach 50 ml, i.e. 400  $\mu$ g/50 ml  $\rightarrow$ 8  $\mu$ g/ml. (Standard). 2 ml from this standard was putted in tube and add 0.1 ml sodium nitrite, vortex and incubate the tube on 22°C for 20 minutes. Then read this solution using spectrophotometer to obtain the optical density of standard.

Precipitation of plasma proteins in plasma samples: 1 ml from each plasma sample was put in one tube and add 1 ml distilled water plus 1 ml zinc reagent plus 1 ml NaOH. Mixing the containing tubes using vortex for  $\frac{1}{2}$  minute and centrifuge the samples tubes on 3500 rpm for 15 minutes to obtain the supernatant.

Estimation of ANP in supernatant of samples: 2 ml from supernatant solution (contains ½ ml plasma) was putted in one tube and add 0.1 ml sodium nitrite and one drop (50 µl)  $\rm H_2SO_4$  (4N) and incubate the tubes on 22°C for 20 minutes. Optical densities of all tubes were reading using Spectrophotometer. Concentration of ANP (µg/ml) in each sample was determined as following: ANP concentration= (Optical density of sample/Optical density of std.) x concentration of standard (8 µg/ml) = µg. Standard tube put in spectrophotometer and optical density of standard was fixed and concentration of ANP in each sample determined directly without the equation.

Estimating of body water: Estimation of body water (ml) in any animal or person is given by concentration of ANP Injected ( $\mu$ g) to concentration of ANP in plasma sample ( $\mu$ g). Body water = ANP injected ( $\mu$ g) / ANP in plasma sample ( $\mu$ g/ml). Estimation was carried out in ½ ml plasma (2 ml from supernatant/4 ml during precipitation of plasma proteins). Therefore multiplied concentration in dilution factor (2) and also multiplied in 100/93 (percentage of water content in plasma) as following:

Body water= [ANP injected ( $\mu g$ )/ ANP in plasma sample ( $\mu g$ / ml)] x 2 x 100/93= liter.

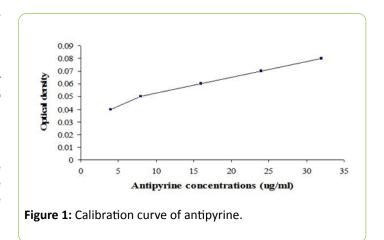
#### Statistical analysis

Data of total body water in five calves by two methods were analyzed statistically using t-paired test according to Snedecor and Cochran [12].

# **Results and Discussion**

# Estimation body water in 5 calves using extrapolation technique

1: In this conventional method, five samples must withdraw after 2 g ANP injection in each calf and make ANP standard curve and extrapolated back to the time of injection by the method of least squares (extrapolation technique). Optical densities are plotted against the corresponding concentrations of ANP ( $\mu$ g/ml) on a semilogaithmic paper as following (Figure 1).



2: Clear supernatant of plasma samples after protein precipitation by centrifugation was added one drop (0.05 ml) of 4 N  $\rm H_2SO_4$  followed by two drops (0.1 ml) of 0.2% sodium solution and then read optical density of samples at different times after dosing (Table 2). From standard curve, the concentrations of ANP in plasma samples ( $\mu \rm g/ml$ ) at different hours after dosing were known as following in Table 3. Plasma levels of antipyrine at various intervals after intravenous injection were plotted on semilogarthmic paper against time in hours.

To correct for the metabolism of the ANP during the time required for uniform distribution, the curve for the plasma level is extrapolated of the logarithm of the plasma concentrations to zero time. The plasma ANP concentration ( $\mu$ g/ml) at zero time is calculated by plotting the plasma levels of ANP (**Figure 2**).

The straight portion of the time concentration curve was extrapolated back to the time of injection (ANP  $\mu g/ml$  at zero time) by the method of least squares. The plasma water level of antipyrine is calculated by dividing the plasma level of antipyrine by the water content of the plasma. The calculation for body water is made as follows: Body water, ml = Amount of ANP injected ( $\mu g$ )/ Amount of ANP in plasma ( $\mu/ml$ ). Body water was estimated as in **Table 4.** 

Estimation body water using modified technique: Estimating body water content in live animal using ANP was carried out by single blood sample at  $\frac{1}{2}$  hour after ANP injection as modified technique in the same calves. Optical density of one sample ( $\frac{1}{2}$  h after injection) and also ANP concentration in one sample at  $\frac{1}{2}$  hour after injection of 2 g ANP in each calf was estimated.

Standard tube put in spectrophotometer and optical density of standard was fixed and concentration of ANP in each sample determined directly according this equation: TBW =  $\{(2 \times 1000 \times 1000)/\text{ANP} \text{ at zero time or at } \frac{1}{2} \text{ hr after dosing} \} \times 2 \times (100/93) = 1 \text{ liter as presented in Table 5. Comparable between convention and modified methods in estimation body water in five calves was in Table 6. Data shows that averages of total body water in five calves were 135.8 and 132.3 liters in extrapolated method and modified method, respectively.$ 

In the present study, average total body water in 5 calves determined by the modified method was 3.5 liters (2.6%) less than that obtained from the extrapolation method. This means

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that the modified method measures about 97.4 of the total body water in calves. However, the values for body water obtained by the two methods did not differ significantly. Because in modified method not depriving the animals of feed and water for 5 h.

In addition, animals do not lose water by vaporization during such a time and their physiological systems are not disturbed by convention method measurement. Besides, modified method is 5 times faster than the convention method. However, Kamal and Habeeb studied the comparison between methods of estimating total body water using tritiated water, Antipyrine and desiccation in Friesian cattle and found that estimating body water using Antipyrine was accurate technique with relation to desiccation method.

**Estimation of heat tolerance coefficient:** Body water is vary considerably in animal during growing due to difference in the rate of accumulation of the less hydrated, fat, collagen and fibrous tissues in replacement of the more hydrated functioning protoplasmic mass [13]. In addition, body water concentration in animal is also differed due to difference in response to nutritional and climatic factors.

Animals when exposed to high ambient temperature during hot summer season, water intake increase and consequently body water content increases with different percentage according animal response to stressful conditions [3]. The percentage change (heat-induced changes) in body water or body solids (Live body weight- body water) contents in each animal may be used for evaluation the animal's adaptability to

heat stress [10]. The heat-induced changes may be used as index for heat tolerance coefficient (HTC).

Detection of such phenomena in the animals could be achieved by body water and body solids as presented. The heat-induced changes in each of total body water and total body solids in live animals by antipyrine dilution technique were used previously as heat tolerance coefficient for detection of heat adaptability in farm animals [3,14,15].

**Estimation of Heat Tolerance Coefficient (HTC) using total body water:** The body water is determined using ANP by modified technique under each of comfortable conditions (BW1) and under heat stress exposure (BW2). The percentage increase in body water due to heat stress conditions may be used as index for heat tolerance coefficient (HTC) as following:

HTC=100-[BW2-BW1/BW1 x 100] where BW1 and BW2 are body water during comfortable and hot conditions, respectively. The most heat tolerance animals are those with the highest values as assuming in **Table 7.** 

Data in **Tables 7 and 8** are assuming values to clear how HTC and adaptability grade were estimated. Habeeb [14] estimated actually this coefficient (HTC) in sheep and goats and concluded that the most heat tolerant animals are those with the highest values.

**Estimation of Heat Tolerance Coefficient (HTC) using total body solids:** It is well known that body weight including body solids and body water. Body solids (BS) = body weight - body water.

Table 2: Optical density of ANP samples after dosing.

| Calf no.  | Time after dosing, hr |       |       |       |  |  |
|-----------|-----------------------|-------|-------|-------|--|--|
| Call IIO. | 1                     | 2     | 3     | 4     |  |  |
| 1         | 0.09                  | 0.08  | 0.072 | 0.07  |  |  |
| 2         | 0.069                 | 0.056 | 0.042 | 0.034 |  |  |
| 3         | 0.073                 | 0.063 | 0.046 | 0.044 |  |  |
| 4         | 0.086                 | 0.066 | 0.042 | 0.032 |  |  |
| 5         | 0.115                 | 0.09  | 0.069 | 0.044 |  |  |

**Table 3:** ANP concentrations (μg/ml) in plasma samples at different hours after dosing from standard curve.

| Calf no.  | Time after dosing , hr |    |    |    |  |  |
|-----------|------------------------|----|----|----|--|--|
| Call IIO. | 1                      | 2  | 3  | 4  |  |  |
| 1         | 29                     | 26 | 23 | 22 |  |  |
| 2         | 22                     | 18 | 14 | 11 |  |  |
| 3         | 24                     | 21 | 15 | 14 |  |  |
| 4         | 27                     | 21 | 14 | 10 |  |  |
| 5         | 37                     | 29 | 23 | 15 |  |  |

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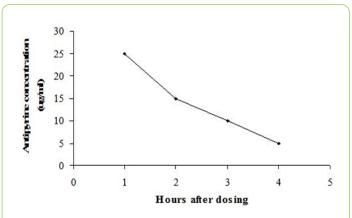


Figure 2: Estimation of antipyrine concentration at zero time.

Estimation of the body water using ANP by modified methods under each of comfortable (BW1) and heat stress (BW2) and each value was subtracted from the corresponding body weight (weight1 and weight2) to obtain body solids under comfortable (BS1) and under heat stress (BS2). Body solids loss due to heat stress may be used as HTC as following:

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HTC =  $100 - [BS2 - BS1 / BS1 \times 100]$ , where BS1 and BS2 are the BS during comfortable and heat stress, respectively, as assuming in Table 8.

Table 4: ANP concentrations at zero time and estimation of body water in five calves using extrapolation technique.

| Calf no. | Body weight of calves, kg | ANP (μg/ml) at zero time | Total body water, liter                                  |
|----------|---------------------------|--------------------------|--|
| 1        | 200                       | 33                       | TBW = { ( 2 x 1000 x 1000 )/33 } x 2 x (100/93) = 130.34 |
| 2        | 230                       | 26.5                     | TBW = { ( 2 x 1000 x 1000 )/26.5} x 2 x (100/93)=162.30  |
| 3        | 235                       | 27                       | TBW = { ( 2 x 1000 x 1000 )/27} x 2 x (100/93) = 159.30  |
| 4        | 185                       | 34                       | TBW = { ( 2 x 1000 x 1000 )/34 } x 2 x(100/93) = 126.50  |
| 5        | 68                        | 44                       | TBW = { (2 x 1000 x 1000)/44} x 2 x (100/93) = 97.75     |

**Table 5:** Estimate total body water in five calves using modified technique.

| Calf no. | O.D. of one sample (½ h after injection) | ANP concentrations in one sample (½ h after injection) | Total body water, liter (Modified technique)             |
|----------|--|--|--|
| 1        | 0.095                                    | 33.5   | TBW ={(2 x 1000 x 1000)/33.5 } x 2 x (100/93) =128.39    |
| 2        | 0.075                                    | 27.5   | TBW ={(2 x 1000 x 1000)/27.5} x 2 x (100/93) = 156.40    |
| 3        | 0.075                                    | 27.5   | TBW = {(2 x 1000 x 1000)/27.5 } x 2 x ( 100/93) = 156.40 |
| 4        | 0.099                                    | 34.5   | TBW = {(2 x 1000 x 1000)/ 34.5} x 2 x (100/93) = 124.67  |
| 5        | 0.131                                    | 45   | TBW = {(2 x 1000 x 1000)/45} x 2 x (100/93) = 95.58      |

Table 6: Estimate body water in calves using convention and modified techniques.

| 0-16     | Body weight of | Convention method |                     | Modified method |                     | Differences |  |
|----------|----------------|-------------------|---------------------|-----------------|---------------------|-------------|--|
| Calf no. | calves, kg     | ANP at 0 time     | Total body water, I | ANP at 0 time   | Total body water. I |             |  |
| 1        | 200            | 33                | 133.34              | .34 33.5 128.39 |                     | -4.95       |  |
| 2        | 230            | 26.5              | 162.3               | 27.5            | 156.4               | -5.9        |  |
| 3        | 235            | 27                | 159.3               | 27.5            | 156.4               | -2.9        |  |
| 4        | 185            | 34                | 126.5               | 34.5            | 124.67              | -1.83       |  |
| 5        | 68             | 44                | 97.75               | 45 95.58        |                     | -2.17       |  |
| Sum      | 918            |                   | 679.2               |                 | 661.4*              | -17.8       |  |
| X ± SE   | 183.6 ± 30.3   | 32.9 ± 3.16       | 135.8 ± 11.8        | 33.6 ± 3.20     | 132.3 ± 11.35       | -3.5        |  |

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| Accuracy %           |  | 97.4 |  |  |
|----------------------|--|------|--|--|
| *NS= Not Significant |  |      |  |  |

Table 7: Estimation of heat tolerance coefficient (HTC) using total body water (BW)

| Animal no.   | BW1 under comfortable conditions | BW2 under heat stress conditions | Change % | *HTC (100-change<br>% ) | Adaptability grade |  |  |
|--|----------------------------------|----------------------------------|----------|-------------------------|--------------------|--|--|
| 1  | 100                              | 120                              | 20       | 80                      | Best               |  |  |
| 2  | 100                              | 130                              | 30       | 70                      | Moderate           |  |  |
| 3  | 100                              | 140                              | 40       | 60                      | Worst              |  |  |
| *Heat tolerance coefficient (HTC) =100 - [BW2 - BW1 / BW1 x 100] |                                  |                                  |          |                         |                    |  |  |

Table 8: Estimation of heat tolerance coefficient (HTC) using total body solids (BS).

| Under comfortable |  | Under heat stress |          |     | Change % in BS | *HTC (100-%<br>change) | Adaptability grade |          |
|-------------------|--|-------------------|----------|-----|----------------|------------------------|--------------------|----------|
| Weight 1          | BW1  | BS1               | Weight 2 | BW2 | BS2            |                        |                    |          |
| 160               | 100  | 60                | 180      | 120 | 60             | 0                      | 100                | Best     |
| 160               | 100  | 60                | 180      | 130 | 50             | 17                     | 83                 | Moderate |
| 160               | 100  | 60                | 180      | 140 | 40             | 33                     | 67                 | Worst    |
| *Heat tolerand    | *Heat tolerance coefficient (HTC) =100 - [BS2 - BS1 / BS1 x 100] |                   |          |     |                |                        |                    |          |

Kamal and Habeeb [4] in Friesian calves and Habeeb and Gad [15] in growing native and crossing bovine calves determined actually this heat tolerance coefficient (HTC) using change in body solids and found that the most heat tolerant animals are those with the highest values.

### Conclusion

It is concluded that estimate body water using ANP by new method is simple, easily, accurate and quickly technique and more reliable and the accuracy of modified technique was 97.4 % as compared with convention method. In addition, the heat-induced changes in each of total body water and total body solids in live animals using antipyrine dilution technique may be used as heat tolerance coefficient for detection of heat adaptability in live animals

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